

Taurine

Carl Henrik Görbitz, Kristian Prydz and Sigurd Ugland

Copyright © International Union of Crystallography

This paper is published electronically. It meets the data-validation criteria for publication in *Acta Crystallographica Section C*. The submission has been checked by a Section C Co-editor though the text in the "Comments" section is the responsibility of the authors.

Taurine

Carl Henrik Görbitz,^{a*} Kristian Prydz^b and Sigurd Ugland^a^aDepartment of Chemistry, University of Oslo, PO Box 1033 Blindern, N-0315 Oslo, Norway, and ^bDepartment of Biochemistry, University of Oslo, PO Box 1041 Blindern, N-0315 Oslo, Norway
Correspondence e-mail: c.h.gorbitz@kjemi.uio.no

Received 1 December 1999

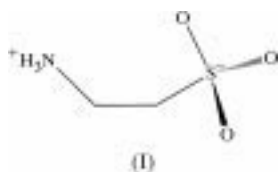
Accepted 2 December 1999

Data validation number: IUC9900178

The structure of taurine (2-aminoethanesulfonic acid), C₂H₇NO₃S, has been redetermined at 150 K, and is compared with the structures of glycine and β -alanine which, like taurine, are enzymatically conjugated with bile acids.

Comment

Bile acids are conjugated with glycine and taurine, (I), to become more water soluble, which facilitates secretion. This enzymatic activity seems to have a bimodal localization in rat liver with different Kms for glycine and taurine, but competition experiments indicate that the same enzymes are able to catalyse conjugation both with glycine and taurine (Kase & Björkhem, 1989). Furthermore, β -alanine (3-aminopropanoic acid, not to be misinterpreted as a second polymorph of \mathcal{L} -alanine) is also a substrate. This makes a structural comparison of these three amino acids interesting. The structure of taurine was first presented by Okaya (1966). We report here a more accurate low-temperature study.



Glycine has been crystallized in three polymorphic forms, α -glycine (Jönsson & Kvick, 1972), β -glycine (Iitaka, 1960) and γ -glycine (Kvick *et al.*, 1980) with different hydrogen-bond patterns. The structure of α -glycine (Jönsson & Kvick, 1972) is very similar to the structure of β -alanine (Jose & Pant, 1965; Papavinasam *et al.*, 1986) in that amino acid dimers (with centers of symmetry) connected by two hydrogen bonds occur in both structures. The corresponding first-level graph-sets (Etter, 1990; Bernstein *et al.*, 1995) are R²₂(10) and R²₂(12), respectively. The conformation at the central C—C bond in β -alanine is *gauche* [N—C—C = 83.2 (2)°, Papavinasam *et al.*, 1986]. A *gauche* orientation is found also for taurine with N1—C2—C1—S1 = 70.96 (5)°. In addition to a weak intra-

molecular contact between the amino- and sulfonic acid groups [graph-set S(6)], the hydrogen-bond pattern again incorporates the centrosymmetric R²₂(12) motif, involving not just one, but two different neighboring molecules resulting in formation of hydrogen-bonded ribbons along the *a* axis. In summary, the structures of glycine (in the α -modification, Jönsson & Kvick, 1972), β -alanine (Papavinasam *et al.*, 1986) and taurine show several common features. In particular, the tendency of these molecules to form hydrogen-bonded ring systems may be important for substrate specificity when conjugation with bile acids occurs.

Experimental

Taurine was purchased from Sigma. The crystals were obtained by diffusion of 2-propanol into 50 μ l of an aqueous solution containing about 2.0 mg of the acid.

Crystal data

C ₂ H ₇ NO ₃ S	$D_x = 1.730 \text{ Mg m}^{-3}$
$M_r = 125.15$	Mo $K\alpha$ radiation
Monoclinic, $P2_1/c$	Cell parameters from 2830 reflections
$a = 5.2729 (1) \text{ \AA}$	$\theta = 3.5\text{--}40.0^\circ$
$b = 11.6565 (3) \text{ \AA}$	$\mu = 0.563 \text{ mm}^{-1}$
$c = 7.8383 (2) \text{ \AA}$	$T = 150 (2) \text{ K}$
$\beta = 94.011 (1)^\circ$	Block, colourless
$V = 480.59 (2) \text{ \AA}^3$	$0.60 \times 0.45 \times 0.40 \text{ mm}$
$Z = 4$	

Data collection

Siemens SMART CCD diffractometer	2891 independent reflections
Sets of exposures each taken over 0.6° ω rotation scans	2736 reflections with $I > 2\sigma(I)$
Absorption correction: empirical (SADABS; Sheldrick, 1996)	$R_{\text{int}} = 0.0273$
$T_{\text{min}} = 0.755$, $T_{\text{max}} = 0.844$	$\theta_{\text{max}} = 40.26^\circ$
7640 measured reflections	$h = -9 \rightarrow 8$
	$k = -21 \rightarrow 20$
	$l = -14 \rightarrow 13$

Refinement

Refinement on F^2	$w = 1/[\sigma^2(F_o^2) + (0.0408P)^2 + 0.0602P]$
$R[F^2 > 2\sigma(F^2)] = 0.026$	where $P = (F_o^2 + 2F_c^2)/3$
$wR(F^2) = 0.075$	$(\Delta/\sigma)_{\text{max}} = 0.010$
$S = 1.126$	$\Delta\rho_{\text{max}} = 0.526 \text{ e \AA}^{-3}$
2891 reflections	$\Delta\rho_{\text{min}} = -0.417 \text{ e \AA}^{-3}$
93 parameters	Extinction correction: <i>SHELXTL</i>
H atoms treated by a mixture of independent and constrained refinement	Extinction coefficient: 0.133 (10)

Table 1
Selected geometric parameters (\AA , °).

S1—O1	1.4543 (5)	S1—C1	1.7842 (6)
S1—O2	1.4690 (5)	N1—C2	1.4911 (7)
S1—O3	1.4699 (4)	C1—C2	1.5249 (8)
O1—S1—C1	106.95 (3)	C2—C1—S1	112.61 (4)
O2—S1—C1	105.71 (3)	N1—C2—C1	112.28 (4)
O3—S1—C1	105.85 (3)		
O1—S1—C1—C2	179.69 (4)	S1—C1—C2—N1	70.96 (5)

Table 2

Hydrogen-bonding geometry (Å, °).

$D-H \cdots A$	$D-H$	$H \cdots A$	$D \cdots A$	$D-H \cdots A$
N1–H1 \cdots O3	0.790 (14)	2.416 (15)	2.924 (1)	123.2 (13)
N1–H1 \cdots O3 ⁱ	0.790 (14)	2.386 (15)	3.000 (1)	135.5 (13)
N1–H1 \cdots O2 ⁱ	0.790 (14)	2.543 (14)	3.218 (1)	144.3 (14)
N1–H2 \cdots O2 ⁱⁱ	0.812 (15)	1.991 (15)	2.789 (1)	167.1 (15)
N1–H3 \cdots O3 ⁱⁱⁱ	0.811 (15)	2.112 (14)	2.879 (1)	157.8 (14)

Symmetry codes: (i) $-x, 1-y, 1-z$; (ii) $-x, \frac{1}{2}+y, \frac{1}{2}-z$; (iii) $1-x, 1-y, 1-z$.

The data collection nominally covered over a hemisphere of reciprocal space by a combination of five sets of exposures; two with the detector set at $2\theta = 30^\circ$ and three with $2\theta = 55^\circ$. Each set had a different φ angle for the crystal and each exposure covered 0.6° in ω . The crystal-to-detector distance was 4.97 cm. Coverage of the unique set is over 99% complete to at least 70° in 2θ . H atoms were refined isotropically.

Data collection: *SMART* (Siemens, 1995); cell refinement: *SAINT* (Siemens, 1995); data reduction: *SAINT* (Siemens, 1995); program(s) used to solve structure: *SHELXTL* (Sheldrick, 1997); program(s) used to refine structure: *SHELXTL* (Sheldrick, 1997); molecular graphics: *SHELXTL* (Sheldrick, 1997); software used to prepare material for publication: *SHELXTL* (Sheldrick, 1997).

The purchase of the Siemens SMART diffractometer was made possible through support from The Research Council of Norway (NFR).

References

- Bernstein, J., Davis, R. E., Shimoni, L., & Chang, N.-L. (1995). *Angew. Chem. Int. Ed. Engl.* **34**, 1555–1573.
- Etter, M. C. (1990). *Acc. Chem. Res.* **23**, 120–126.
- Iitaka, Y. (1960). *Acta Cryst.* **13**, 35–45.
- Jönsson, P.-G. & Kvik, Å. (1972). *Acta Cryst.* **B28**, 1827–1833.
- Jose, P. & Pant, L. M. (1965). *Acta Cryst.* **18**, 806–810.
- Kase, B. F. & Björkhem, I. (1989). *J. Biol. Chem.* **264**, 9220–9223.
- Kvik, Å., Canning, W. M., Koetzle, T. F. & Williams, G. J. B. (1980). *Acta Cryst.* **B36**, 115–120.
- Okaya, Y. (1966). *Acta Cryst.* **21**, 726–735.
- Papavinasam, E., Natarajan, S. & Shivaprakash, N. C. (1986). *Int. J. Pept. Protein Res.* **28**, 525–527.
- Sheldrick, G. M. (1996). *SADABS*. University of Göttingen, Germany.
- Sheldrick, G. M. (1997). *SHELXTL*. Version 5.0. Siemens Analytical X-ray Instruments Inc., Madison, Wisconsin, USA.
- Siemens (1995). *SMART* and *SAINT*. Siemens Analytical X-ray Instruments Inc., Madison, Wisconsin, USA.